



BD BBL™ Phenylethyl Alcohol Agar with 5% Sheep Blood

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QUALITY CONTROL PROCEDURES

I. INTRODUCTION

Phenylethyl Alcohol Agar with 5% Sheep Blood is a selective medium for the isolation of gram-positive organisms from clinical and nonclinical materials.

II. PERFORMANCE TEST PROCEDURE

- Inoculate representative samples with the cultures listed below.
 - Using a volumetric pipettor or equivalent method, deliver 0.1 mL of a dilution yielding 30 - 300 CFU to each plate and spread-inoculate using a sterile glass spreader.
 - Incubate the plates at $35 \pm 2^\circ\text{C}$ in an aerobic atmosphere supplemented with 3-5% carbon dioxide.
 - Include **Trypticase** Soy Agar with 5% Sheep Blood (TSA5%SB) plates as nonselective controls for all organisms.
- Examine plates after 18-24 h for growth, colony size, hemolytic reactions and selectivity.
- Expected Results

CLSI Organisms	ATCC™	Recovery
* <i>Streptococcus pyogenes</i>	19615	Growth
* <i>Staphylococcus aureus</i>	25923	Growth
* <i>Proteus mirabilis</i>	12453	Inhibition (partial)

Additional Organism

<i>Streptococcus pneumoniae</i>	6305	Growth
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*Recommended organism strain for User Quality Control.

III. ADDITIONAL QUALITY CONTROL

- Examine plates as described under "Product Deterioration."
- Visually examine representative plates to assure that any existing physical defects will not interfere with use.
- Determine the pH potentiometrically at room temperature for adherence to the specification of 7.3 ± 0.2 .
- Note the firmness of plates during the inoculation procedure.
- Incubate uninoculated representative plates at $30 \pm 1^\circ\text{C}$ for 84 h and examine for microbial contamination.

PRODUCT INFORMATION

IV. INTENDED USE

Phenylethyl Alcohol Agar with 5% Sheep Blood is a selective medium for the isolation of gram-positive organisms, particularly gram-positive cocci, from specimens of mixed gram-positive and gram-negative flora. The medium should not be used for determination of hemolytic reactions since atypical reactions may be observed.

V. SUMMARY AND EXPLANATION

After having noted that phenylethyl alcohol exhibited a marked inhibitory effect on gram-negative bacteria with only slight effect on gram-positive organisms, Lilley and Brewer incorporated the chemical in an infusion agar base as a selective agent for the isolation of gram-positive bacteria.¹ Phenylethyl Alcohol with 5% Sheep Blood is used in the microbiology laboratory to inhibit gram-negative bacteria, particularly *Proteus*, in specimens containing a mixed bacterial flora.

VI. PRINCIPLES OF THE PROCEDURE

Phenylethyl Alcohol Agar with 5% Sheep Blood supports the growth of gram-positive bacterial species, due

to its content of peptones, which supply nitrogen, carbon, sulfur and trace nutrients. Sodium chloride maintains osmotic equilibrium. Sheep blood is a source of many growth factors. Phenylethyl alcohol is bacteriostatic for gram-negative bacteria since it selectively and reversibly inhibits DNA synthesis.²

VII. REAGENTS

Phenylethyl Alcohol Agar with 5% Sheep Blood

Approximate Formula* Per Liter Purified Water

Pancreatic Digest of Casein	15.0 g
Papaic Digest of Soybean Meal	5.0 g
Sodium Chloride	5.0 g
β-Phenylethyl Alcohol	2.5 g
Agar	15.0 g
Sheep Blood, defibrinated	5%

*Adjusted and/or supplemented as required to meet performance criteria.

Warnings and Precautions:

For *in vitro* Diagnostic Use in Taiwan

If excessive moisture is observed, invert the bottom over an off-set lid and allow to air dry in order to prevent formation of a seal between the top and bottom of the plate during incubation.

Pathogenic microorganisms, including hepatitis viruses and Human Immunodeficiency Virus, may be present in clinical specimens. "Standard Precautions"³⁻⁶ and institutional guidelines should be followed in handling all items contaminated with blood and other body fluids. After use, prepared plates, specimen containers and other contaminated materials must be sterilized by autoclaving before discarding.

Storage Instructions: On receipt, store plates in the dark at 2-8°C. Avoid freezing and overheating. Do not open until ready to use. Minimize exposure to light. Prepared plates stored in their original sleeve wrapping at 2-8°C until just prior to use may be inoculated up to the expiration date and incubated for recommended incubation times. Allow the medium to warm to room temperature before inoculation.

Product Deterioration: Do not use plates if they show evidence of microbial contamination, discoloration, drying, cracking or other signs of deterioration.

VIII. SPECIMEN COLLECTION AND HANDLING

Specimens suitable for culture may be handled using various techniques. For detailed information, consult appropriate texts.^{7,8}

Specimens should be obtained before antimicrobial therapy has been administered. Provision must be made for prompt delivery to the laboratory.

IX. PROCEDURE

Material Provided: Phenylethyl Alcohol Agar with 5% Sheep Blood

Materials Required But Not Provided: Ancillary culture media, reagents, quality control organisms and laboratory equipment as required.

Test Procedure: Observe aseptic techniques.

The agar surface should be smooth and moist, but without excessive moisture.

Streak the specimen as soon as possible after it is received in the laboratory. The streak plate is used primarily to isolate pure cultures from specimens containing mixed flora. Alternatively, if material is being cultured directly from a swab, roll the swab over a small area of the surface at the edge and streak from this inoculated area.

Incubate plates 24-48 h at 35 ± 2°C in an aerobic atmosphere supplemented with carbon dioxide.

User Quality Control: See "Quality Control Procedures."

Quality Control requirements must be performed in accordance with applicable local, state and/or federal regulations or accreditation requirements and your laboratory's standard Quality Control procedures. It is recommended that the user refer to pertinent CLSI (formerly NCCLS) guidance and CLIA regulations for appropriate Quality Control practices.

X. RESULTS

After incubation, most plates will show an area of confluent growth. Because the streaking procedure is, in effect, a "dilution" technique, diminishing numbers of microorganisms are deposited on the streaked areas. Consequently, one or more of these areas should exhibit isolated colonies of the organisms contained in the

specimen. Better isolation is obtained due to the inhibitory action of the medium.

Typical colonial morphology on Phenylethyl Alcohol Agar with 5% Sheep Blood is as follows.

Streptococci. Small, white to gray. May exhibit alpha or beta hemolysis but hemolysis may be atypical.

Enterococci (group D). Small, but larger than group A streptococci, and blue-gray. May exhibit alpha or beta hemolysis.

Staphylococci. Large, white to gray or cream to yellow. Hemolytic reactions are variable.

Micrococci Large, white to gray or yellow to orange. Hemolytic reactions are variable.

Corynebacteria Small to large, white to gray to yellow. Hemolytic reactions are variable.

Candida Small, white

Listeria monocytogenes Small to large, blue-gray and beta-hemolytic

Gram-negative bacteria No growth to moderate growth; swarming inhibited.

XI. LIMITATIONS OF THE PROCEDURE

For identification, organisms must be in pure culture. Morphological, biochemical and/or serological tests should be performed for final identification. Consult appropriate texts for detailed information and recommended procedures.⁷⁻¹²

A single medium is rarely adequate for detecting all organisms of potential significance in a specimen. It should be recognized that organisms generally susceptible to the antimicrobial agent in a selective medium may be completely or only partially inhibited depending upon the concentration of the agent, the characteristics of the microbial strain and the number of organisms in the inoculum. Organisms that are generally resistant to the antimicrobial agent should not be inhibited. Cultures of specimens grown on selective media should, therefore, be compared with specimens cultured on nonselective media to obtain additional information and help ensure recovery of potential pathogens.

XII. AVAILABILITY

Cat. No.	Description
212086	BD BBL™ Phenylethyl Alcohol Agar with 5% Sheep Blood, Pkg. of 20 plates

XIII. REFERENCES

1. Lilley, B.D., and J.H. Brewer. 1953. The selective antibacterial action of phenylethyl alcohol. *J. Am. Pharm. Assoc.* 42:6-8.
2. Dowell, V.R., Jr., E.O. Hill, and W.A. Altemeier. 1964. Use of phenylethyl alcohol in media for isolation of anaerobic bacteria. *J. Bacteriol.* 88:1811-1813.
3. Clinical and laboratory Standards Institute. 2005. Approved Guideline M29-A3. Protection of laboratory workers from occupationally acquired infections, 3rd ed. CLSI, Wayne, PA.
4. Garner, J.S. 1996. Hospital Infection Control Practices Advisory Committee, U.S. Department of Health and Human Services, Centers for Disease Control and Prevention. Guideline for isolation precautions in hospitals. *Infect. Control Hospital Epidemiol.* 17:53-80.
5. U.S. Department of Health and Human Services. 1999. Biosafety in microbiological and biomedical laboratories, HHS Publication (CDC), 4th ed. U.S. Government Printing Office, Washington, D.C.
6. Directive 2000/54/EC of the European Parliament and of the Council of 18 September 2000 on the protection of workers from risks related to exposure to biological agents at work (seventh individual directive within the meaning of Article 16(1) of Directive 89/391/EEC). *Official Journal L262*, 17/10/2000, p. 0021-0045.
7. Murray, P.R., E.J. Baron, J.H. Jorgensen, M.A. Pfaller, and R. H. Tenover (ed.). 2003. *Manual of clinical microbiology*, 8th ed. American Society for Microbiology, Washington, D.C.
8. Forbes, B.A., D.F. Sahm, and A.S. Weissfeld. 2002. *Bailey and Scott's diagnostic microbiology*, 11th ed. Mosby, Inc., St. Louis.
9. Holt, J.G., N.R. Krieg, P.H.A. Sneath, J.T. Staley, and S.T. Williams (ed.). 1994. *Bergey's Manual of determinative bacteriology*, 9th ed. Williams & Wilkins, Baltimore.
10. MacFaddin, J.F. 2000. *Biochemical tests for identification of medical bacteria*, 3rd ed. Lippincott Williams & Wilkins, Baltimore.
11. Koneman, E.W., S.D. Allen, W.M. Janda, P.C. Schreckenberger, and W.C. Winn, Jr. 1997. *Color atlas and textbook of diagnostic microbiology*, 5th ed. Lippincott-Raven, Philadelphia.
12. Isenberg, H.D. (ed.). 2004. *Clinical microbiology procedures handbook*, vol. 1, 2 and 3, 2nd ed. American Society for Microbiology, Washington, D.C.

XIV. FURTHER INFORMATION

For further information please contact your local BD representative.

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